



Stereoselective Synthesis of 2',3'-Dideoxy-nucleosides via Intramolecular Glycosylation of Phenyl 1-Seleno-glycosides. Synthesis of 2',3'-Dideoxythymidine.

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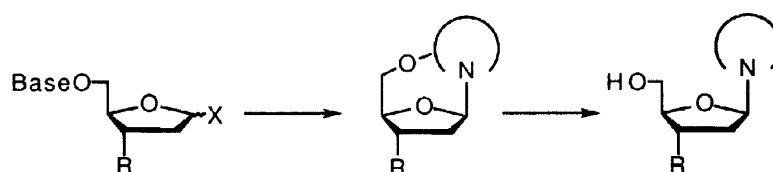
Abstract: 4-methoxy and 4-(2-trimethylsilylethoxy)pyrimidine bases were attached to the 5-position of the phenyl 2,3-dideoxy-1-seleno-*glycero*-pentofuranoside moiety. The presence of the silyl protecting group in the base is necessary to lead to neutral β -anhydro nucleosides by intramolecular glycosylation. The subsequent ring opening affords 3'-deoxythymidine with complete stereocontrol. © 1998 Elsevier Science Ltd. All rights reserved.

Since the discovery of the antiviral activity of 3'-azido-3'-deoxythymidine (AZT)¹ against the human immunodeficiency virus (HIV), there has been intense concern with synthesizing 2',3'-dideoxynucleosides and their analogs² as potential antiviral and antibiotic agents.

Although the standard Vorbrüggen couplings³ between 2-deoxyribose derivatives and pyrimidine or purine bases is one of the simplest methods for obtaining such nucleoside derivatives, the main problem in this approach is the lack of stereocontrol. Moreover, only β -nucleosides usually exhibit high biological activity.

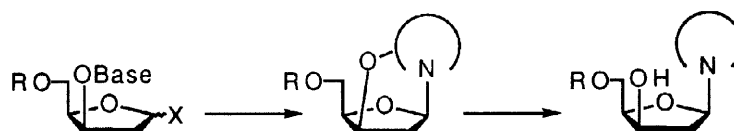
An attractive way of forming selectively β -nucleosides is based on the intramolecular glycosylation strategy. The key step in this approach is the attachment of the base to a 2-deoxy-pentofuranoside at the 5-^{4,5,6} (Scheme 1) or 3- β -position⁷ (Scheme 2), and the formation of a β -anhydro-nucleoside intermediate by intramolecular attack on C-1. Lastly, a β -elimination⁴ at C-5/C-6 or a basic hydrolysis⁵⁻⁷ leads to the desired nucleoside.

Scheme 1



X= OMe	R= OTBDPS	Ref. 4
X= SPh	R=OBn, H, N ₃ , F	Ref. 5
X= OAc	R= OMe	Ref. 6

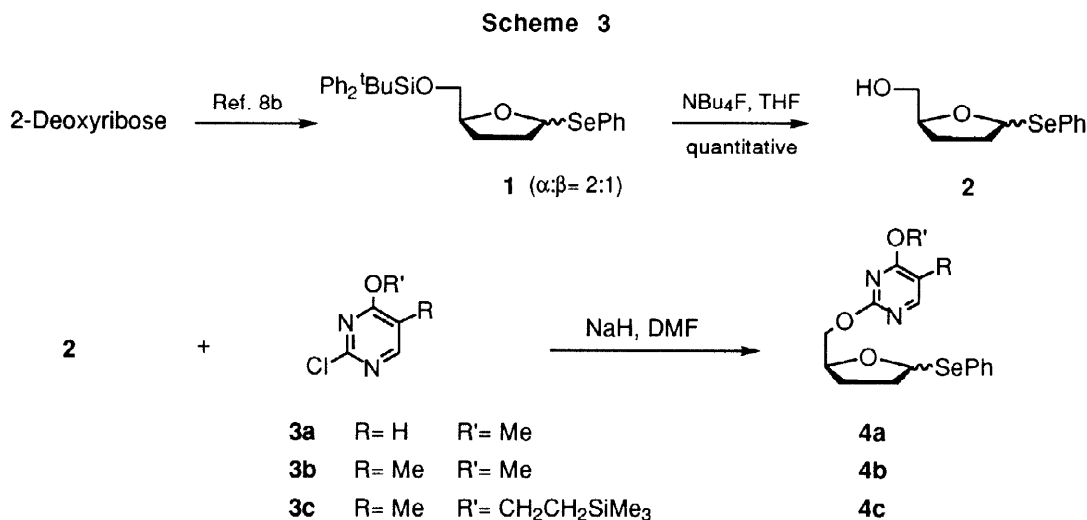
Scheme 2



X = OBn	R = Tr, MOM	Ref. 7
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Hence, as part of a general project which aims to use selenium in the stereoselective synthesis of 2'-deoxy and 2',3'-dideoxy-nucleosides,⁸ we devised an intramolecular strategy to synthesise them from selenoglycosides.

Thus, the starting selenoglycoside **2**⁹ was prepared from 2-deoxyribose in five steps (Scheme 3): methyl glycoside synthesis, selective 5-OH protection, Barton deoxygenation, treatment with PhSeH in the presence of BF₃·OEt₂ to give compound **1**^{8b} and deprotection of the ^tBuPh₂Si group.



On the other hand, pyrimidine derivatives (**3a**,^{10a} **3b**^{10b} and **3c**¹¹) were prepared from 2,4-dichloropyrimidine and 2,4-dichloro-5-methylpyrimidine. Subsequently, **3a**, **3b** and **3c** were attached to phenyl 1-seleno-glycoside **2** following reported procedures;^{5,7} using sodium hydride in DMF compounds **4a**, **4b** and **4c**¹² were obtained in 60-70% yields (Scheme 3).

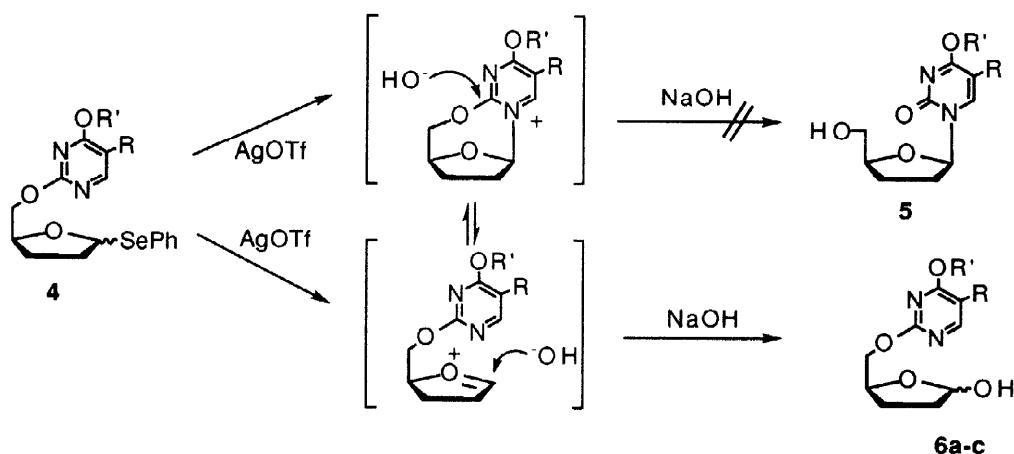
We initially explored the intramolecular glycosylation from **4a** and **4b**. The starting material generated –by activating the anomeric position with AgOTf– a charged anhydronucleoside in equilibrium with the oxonium ion (Scheme 4). The subsequent hydrolysis will lead to the formation of the desired nucleoside **5** or the furanose **6**.

However, all the attempts of glycosylation from the selenoglycosides **4a** and **4b** were negative. The reaction with AgOTf was monitored by TLC which indicated that the starting material disappeared and a different product with lower R_f was formed. But the single product isolated from the subsequent hydrolysis with 1N-NaOH at 0°C⁵ was the C-1 hydrolyzed products **6a** and **6b** respectively, in quantitative yield (Scheme 4). The same product was obtained when the hydrolysis was carried out with a saturated solution of Na₂CO₃. No glycosylation products such as **5** were observed.

In view of these results we next turned our attention to the use of the silyl-protected product **4c**. The idea was to generate a charged intermediate such as **7** which could attack at the C-1 after anomeric group had been activated by the silver salt (Scheme 5). The formation of a neutral β-anhydronucleoside would prevent the problem of hydrolysis, since it is well known that this kind of product reacts selectively at the 2-position of the base under hydrolytic conditions.

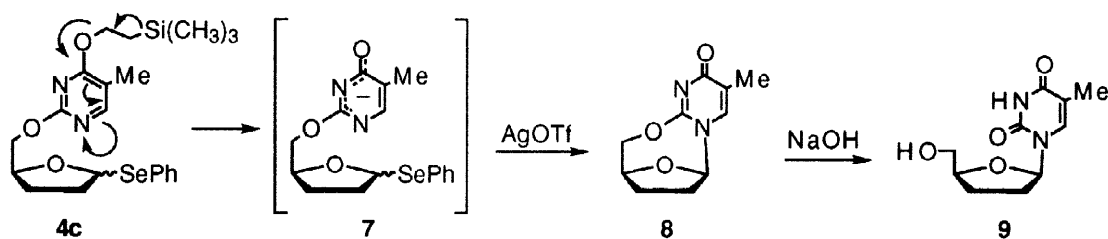
By treating **4c** with AgOTf under the usual conditions (4Å molecular sieves, anhydrous CH₃CN, -20°C, and then NaOH 1N, 0°C) the furanose **6c** (R=Me, R'=CH₂CH₂SiMe₃) was obtained. We also tried to remove the silyl protecting group at the same time activating the anomeric position with AgF. In this case, only the starting material was recovered and no reaction occurred. We also used other reagents such as KF/crown ether or F₂HK used in conjunction with AgOTf and these too gave negative results. The problem was ultimately solved

Scheme 4



by treating **4c** with Bu₄NF in anhydrous CH₃CN at r.t. and then adding AgOTf. Thus, the corresponding 3'-deoxy-2,5'-anhydro-thymidine **8**^{13,14} was obtained in 68% yield. Minor quantities of hydrolyzed product **6** were also recovered. Finally, basic hydrolysis of **8** led to 1-(2,3-dideoxy-β-D-glycero-pentofuranosyl)thymine **9**¹⁵ in quantitative yield.

Scheme 5



In conclusion, 3'-deoxythymidine was synthesized from a phenyl 1-seleno-glycoside via intramolecular glycosylation. The key step is the deprotection of the silyl group at the 4-position of the pyrimidine ring using Bu₄NF prior to activation of the selenoglycoside, and the subsequent formation of a neutral β-anhydronucleoside.

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References and Notes

- Mitsuya, H.; Weinhold, K.J.; Furman, P.A.; St. Clair, M.H.; Lehrman, S.N.; Gallo, R.C.; Bolognesi, D.; Barry, D.W.; Broder, S. *Proc. Natl. Acad. Sci. U.S.A* **1985**, *82*, 7096.
- For reviews see, a) Dueholm, K.L.; Pedersen, E.B. *Synthesis* **1992**, *1*. b) Huryn, D.M.; Okabe, M. *Chem. Rev.* **1992**, *92*, 1745.
- a) Niedballa, U.; Vorbrüggen, H. *J. Org. Chem.* **1974**, *39*, 3654, 3660, 3664. b) Vorbrüggen, H.; Bennua, B. *Chem. Ber.* **1981**, *114*, 1279, and references therein.
- a) Lipshutz, B.H.; Hayakawa, H.; Kato, K.; Lowe, R.F.; Stevens, K.L. *Synthesis* **1994**, 1476. b) Lipschutz, B.H.; Stevens, K.L.; Lowe, R.F. *Tetrahedron Lett.* **1995**, *36*, 2711.

5. a) Sujino, K.; Sugimura, H. *Chemistry Lett.* **1993**, 1187. b) Sujino, K.; Sugimura, H. *Tetrahedron Lett.* **1994**, 35, 1883.
6. Jung, M.E.; Castro, C. *J. Org. Chem.* **1993**, 58, 807.
7. Xia, X., Wang, J., Hager, M.W., Sisti, N., Liotta, D.C. *Tetrahedron Lett.* **1997**, 38, 1111.
8. a) El-Laghdach, A.; DÍaz, Y.; Castellón, S. *Tetrahedron Lett.* **1993**, 34, 2821. b) DÍaz, Y.; El-Laghdach, A.; Matheu, M.I.; Castellón, S. *J. Org. Chem.* **1997**, 62, 1501. c) DÍaz, Y.; El-Laghdach, A.; Castellón, S. *Tetrahedron* **1997**, 53, 10921.
9. (2 α): ^1H NMR (300 MHz, CDCl_3) δ 7.63-7.25 (m, 5H, Ph), 5.94 (dd, 1H, $J_{1,2a}$ = 6.9 Hz, $J_{1,2b}$ = 3.3 Hz, H-1), 4.33 (m, 1H, H-4), 3.75 (dd, 1H, $J_{5',5''}$ = 11.9 Hz, $J_{5,4}$ = 5.9 Hz, H-5), 3.54 (dd, 1H, $J_{5',4}$ = 6.7 Hz, H-5'), 2.46-1.67 (m, 4H, H-2, H-3). ^{13}C NMR (75.4 MHz, CDCl_3) δ 133.9-127.3 (Ph), 84.6 (C-1), 79.2 (C-4), 63.8 (C-5), 33.8 (C-2), 25.8 (C-3). Anal. Calcd for $\text{C}_{11}\text{H}_{14}\text{O}_2\text{Se}$: C, 50.76; H, 5.49. Found: C, 51.05; H, 5.49.
(2 β): ^1H NMR (300 MHz, CDCl_3) δ 7.60-7.20 (m, 5H, Ph), 5.90 (dd, 1H, $J_{1,2a}$ = 6.7 Hz, $J_{1,2b}$ = 2.8 Hz, H-1), 4.30 (m, 1H, H-4), 3.8 (m, 2H, H-5, H-5'), 2.50-1.50 (m, 4H, H-2, H-3). ^{13}C NMR (75.4 MHz, CDCl_3) δ 134.0-127.4 (Ph), 84.6 (C-1), 82.6 (C-4), 64.2 (C-5), 34.6 (C-2), 26.2 (C-3).
10. a) Pluskota, D.; Jankowski, A.; Koroniak, H. *Synth. Commun.* **1992**, 22, 2927. b) Katritzky, A.R.; Baykunt, G.; Rachwal, S.; Szafran, M.; Caster, K.C.; Eyler, J. *J. Chem. Soc., Perkin Trans. 2* **1989**, 1499.
11. 2-Chloro-5-methyl-4-(2'-(trimethylsilyl)ethoxy)pyrimidine **3c**: A solution of 2,4-dichloro-5-methylpyrimidine¹⁰ (12.3 mmol) in THF (6mL) was cooled to -20°C and a mixture of sodium 2-(trimethylsilyl)ethoxide (1.2 mol) in THF (6.5 mL) was added dropwise. The sodium 2-(trimethylsilyl)ethoxide was prepared from 2-(trimethylsilyl)ethanol and NaH. After a night at -20°C , workup of the reaction afforded a crude that was chromatographed over silica gel, eluting with EtOAc/hexane = 1:6. The residue obtained was further purified by means of radial chromatography to give compound **3c** as a white solid (82%); mp: 51-52 $^\circ\text{C}$. ^1H NMR (300 MHz, CDCl_3) δ 8.05 (s, 1H, H-6), 4.46 (t, 2H, $J_{\text{CH}_2\text{-O,CH}_2\text{-Si}}$ = 8.2 Hz, $\text{CH}_2\text{-O}$), 2.07 (s, 3H, CH_3), 1.11 (t, 2H, $\text{CH}_2\text{-Si}$), 0.06 (s, 9H, $\text{Si}(\text{CH}_3)_3$). ^{13}C NMR (75.4 MHz, CDCl_3) δ 168.9 (C-4), 157.3 (C-6), 116.7 (C-5), 65.8 ($\text{CH}_2\text{-O}$), 17.2 ($\text{CH}_2\text{-Si}$), -1.4 ($\text{Si}(\text{CH}_3)_3$). Anal. Calcd for $\text{C}_{10}\text{H}_{17}\text{N}_2\text{OSiCl}$: C, 49.38; N, 11.42; H, 7.26. Found: C, 49.16; N, 11.56; H, 7.10.
12. Spectroscopic data of compounds **4c α** , and **4c β** :
(**4c α**): ^1H NMR (300 MHz, CDCl_3) δ 7.94 (d, 1H, J = 0.9 Hz, H-6'), 7.64-7.24 (m, 5H, SePh), 6.04 (dd, 1H, $J_{1,2A}$ = 6.9 Hz, $J_{1,2B}$ = 2.8 Hz, H-1), 4.66 (ddd, 1H, H-4), 4.47 (t, 2H, $J_{\text{CH}_2\text{-O,CH}_2\text{-Si}}$ = 8.1 Hz, $\text{CH}_2\text{-O}$), 4.39 (dd, 2H, H-5A, H-5B), 2.60-1.80 (m, 4H, H-3, H-2), 1.12 (t, 2H, $\text{CH}_2\text{-Si}$), 0.07 (s, 9H, $\text{Si}(\text{CH}_3)_3$). ^{13}C NMR (75.4 MHz, CDCl_3) δ 169.3 (C-4'), 163.4 (C-2'), 156.7 (C-6'), 133.7-127.2 (SePh), 111.1 (C-5'), 84.70 (C-1), 76.8 (C-4), 68.1 (C-5), 64.7 ($\text{CH}_2\text{-O}$), 33.6 (C-2), 27.2 (C-3), 17.3 ($\text{CH}_2\text{-Si}$), 12.9 (CH_3), -1.4 ($\text{Si}(\text{CH}_3)_3$).
(**4c β**): ^1H NMR (300 MHz, CDCl_3) δ 7.96 (d, 1H, J = 0.9 Hz, H-6'), 7.63-7.24 (m, 5H, SePh), 5.88 (dd, 1H, $J_{1,2A}$ = 6.4 Hz, $J_{1,2B}$ = 3.2 Hz, H-1), 4.52 (ddd, 1H, H-4), 4.47 (t, 2H, $J_{\text{CH}_2\text{-O,CH}_2\text{-Si}}$ = 8.2 Hz, $\text{CH}_2\text{-O}$), 4.38 (dd, 2H, H-5A, H-5B), 2.50-2.10 (m, 4H, H-3, H-2), 2.04 (d, 3H, J = 0.8 Hz, CH_3), 1.13 (t, 2H, $\text{CH}_2\text{-Si}$), 0.08 (s, 9H, $\text{Si}(\text{CH}_3)_3$). ^{13}C NMR (75.4 MHz, CDCl_3) δ 169.3 (C-4'), 163.3 (C-2'), 156.7 (C-6'), 134.2-127.3 (SePh), 111.1 (C-5'), 84.00 (C-1), 79.3 (C-4), 69.3 (C-5), 64.7 ($\text{CH}_2\text{-O}$), 34.1 (C-2), 28.3 (C-3), 17.3 ($\text{CH}_2\text{-Si}$), 12.1 (CH_3), -1.3 ($\text{Si}(\text{CH}_3)_3$).
13. a) Lin, T-S.; Shen, Z-Y.; August, E.M.; Brankovan, V.; Yang, H.; Ghazzouli, I.; Prusoff, W.H. *J. Med. Chem.* **1989**, 32, 1891. b) For the related 2,5'-anhydrothymidine see Watanabe, K.A.; Reichman, U.; Chu, C.K.; Fox, J.J. *Nucl. Acid. Chem.* vol 1, Tipson, R.S.; Townsend, L.B. eds.; John Wiley and Sons, New York,; **1978**, p. 273.
14. **8**: m.p.: 199-201 $^\circ$ (reported 202-203)¹³; UV (MeOH) λ_{max} 248 nm; IR (ν max in cm^{-1}) 1639, 1529, 1474, 1285, 1080; ^1H NMR (300 MHz, CD_3OD) 7.35 (s, 1H, H-6), 5.62 (d, 1H, 7.1 Hz, H-1'), 4.56 (d, 1H, 7.4 Hz, H-4'), 4.22 (d, 1H, 12.0 Hz, H-5'), 4.03 (d, 1H, 12 Hz, H-5''), 2.42 (m, 1H, H-2'), 2.04 (m, 3H, H-2'', H-3', H-3''), 1.82 (s, 3H, Me). ^{13}C NMR (75.4 MHz, CD_3OD) 172.95 (C-4), 157.57 (C-2), 139.01 (C-6), 118.60 (C-5), 94.39 (C-1'), 79.02 (C-4'), 76.82 (C-5'), 31.97 (C-2'), 26.10 (C-3'), 12.45 (Me). Elemental Analysis for $\text{C}_{10}\text{H}_{12}\text{N}_2\text{O}_3$: Calcd. C 57.69, H 5.77, N 13.46. Found C 57.40, H 5.71, N 13.30.
15. Spectroscopic data of compound **9** (^1H and ^{13}C) agrees with a that of a commercial sample and with the previously reported, Barren Beach, J.; Kim, H.O.; Jeong, L.S.; Nampalli, S.; Islam, Q.; Ahn, S.K.; Babu, J.R.; Chu, C.K. *J. Org. Chem.* **1992**, 57, 3887.